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(54) A novel diagnostic marker for splicing variants of genes associated with neurological function

(57) Methods are described for detecting the presence or absence of a four amino acid motif (VRXQ) in expressed proteins that arise from aberrant alternative splicing of premRNA in genes associated with normal neurological function which are useful for detecting neurodegenerative disease. The presence of these variants suggest that mutational events in these genes have occurred. Methods to measure the levels of gene expression of such genes to detect neurodegenerative diseas are provided. Nucleotide sequences and intron-exon junctional sequences of examples of this splicing variant and probes for detecting this variant which are useful as diagnostic reagents are also provided.

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Description

BACKGROUND OF THE INVENTION

In eukaryotes, the initial transcription of genomic DNA into RNA proceeds in the nucleus and yields a contiguous full-length reverse complementary heteronuclear RNA (hnRNA) primary transcript. The hnRNA contains regions or contiguous blocks of nucleotide sequence that end up in the final mRNA (exons) interspersed between "intervening" nucleotide sequences (introns) that do not. In addition to adenylyl methylation and polyadenylation, these hnRNAs are extensively modified in a process referred to as RNA "splicing" wherein discontiguous exons are joined and the intervening intron precisely deleted as an RNA "lariat" from the final mature mRNA transcript (B. Rushkin et al. Cell 1984, 38:317; R.A. Padgett et al. Science 1984, 225:898). RNA splicing is a complex process involving large protein-RNA assemblies called spliceosomes that coordinate the concerted excision and ligation events to yield intron-free mRNAs (M.M. Konarska and P.A. Sharp Cell 1987, 49:763; R. Reid et al. Cell 1988, 53:949; T.A. Steitz Sci. Am. 1988, 258:56).

In normal RNA processing, the resultant mRNA reflects the linear sequence orientation of the exons in the hnRNA; however all exons do not end up in the final transcripts. Rather, several of the resultant mRNAs have only certain exons that result from "alternatively spliced" hnRNA, wherein discontiguous intron-exon junctions are spliced to bring for instance exon1 and exon 4 into juxtaposition rather than exon1 and exon 2. Therefore, several mRNAs may arise from one gene sequence or hnRNA. Not all possible combinations of exons are normally represented in actual mRNA pools arising from one hnRNA as determined by mRNA, cDNA and protein analyses. As an example with three exons (Figure 1), while seven combinations are possible (exon1-exon2-exon3, exon1-exon2, exon1-exon3, exon2-exon3, exon1, exon2, or exon3) perhaps only two (exon1-exon2-exon3 and exon1-exon3) may actually result and be expressed at any appreciable level. These alternatively spliced transcripts are sometimes referred to as "variants". However, for purposes of this invention splice "variant" refers to heretofore unrepresented or expressed mRNAs arising from potential alternative splice sites that result from genomic mutation altering the structure of the hnRNA so that these splices now occur.

The location of splice sites in an hnRNA primary transcript can be determined by comparing the sequences of th corresponding genomic DNA with that of cDNA prepared by copying the corresponding mature mRNA. Any discontinuities between the genomic DNA and cDNA sequences mark the exon-intron boundaries. Such analyses of a number of different RNAs have defined moderately -short "consensus" sequences at the intron-exon boundaries in pre-mRNA and a tendency for a pyrimidine-rich region just upstream of the 3' splice junction (Figure 2). The only universally conserved nucleotides are the first two (GU) and last two (AG) in the intron (Figure 2), though there is a propensity for AG at the 5' exon termini and an initial G at the 3' exon. Only 30-40 nucleotides in the center portion of introns ar necessary for efficient splicing. There is also a conserved A within the context of the pyrimidine rich region of the intron (Figure 2) (..PyrPyrPurAPyrnAG; where Pyr is a pyrimidine and Pur is a purine nucleotide) which is the branch point where the cleaved 5' exon-intron junction loops back to form the "lariat" splicing intermediate (Padgett et al. Science 1984, 225:898). Genetic point mutations that delete or alter these conserved intronic nucleotides (5' GU, 3' AG, or branch point A) would eliminate these splice junctions and prevent normal splicing yielding aberrantly truncated transcripts or transcripts where this exon is deleted and another downstream exon spliced in, that normally may not be spliced in.

A final mechanism for splice variation occurs when several GU or AG dinucleotide motifs occur near consensus intron splice regions of 5' exon-intron or 3' intron-exon boundaries, respectively, such that the splicing system may sometimes not correctly distinguish the correct splice site resulting in alternate protein product some of which may be non-functional or aberrant.

Multiple examples of splice variations exist, many of which are associated with diseases or related disorders. Previous genetic linkage studies have shown a G to A mutation at the 3' splice junction of exon 8 of the gene encoding lysosomal acid lipase. Defects in this gene are associated with cholesterol ester storage disease that result in premature artherosclerosis, hepatomegaly, and elevated LDL cholesterol (U. Seedorf et al. Arterioscler. Throb. Vasc. Biol. 1995, 15: 773-778). Two mutations at the exon 1/intron 1 boundary altered the hepatic specific splicing of the human hydroxymethylbilane synthase gene (third enzyme in heme biosynthetic pathway) and resulted in an enzyme with half-normal activity (K.H. Astrin Human Mutat. 1994, 4:243-252). Deficiency of this enzyme activity eventually results in acute intermittent porphyria (AIP), an autosomal dominant inborn error of metabolism in which life-threatening attacks are precipitated by ecog netic factors. Molecular cloning of cDNA and g nomic DNA hav provided probes allowing presymptomatic detection of these gene defects. In M nk 's disease, a point mutation at the - 2 exonic position of a splice donor site in th middle of the gene causes exon-skipping and activation of a cryptic splice acceptor site (S.G. Kaler et al. Nat. Genet. 1994, 8:195-202). Exon skipping of the entire exon 19 results from a G to A point mutation at the 5' donor site of intron 19 in muscle phosphofructokinase deficiency (T. Hamaguchi Biochem. Biophys. Res. Comm. 1994, 202:444-449). Aberrant RNA splicing from a splice site mutant in the interleukin-2 receptor gamma (gIL2-R) gene results in the generation of an abundant non-functional gIL2-R containing a small intronic insertion and a second

mutant form with 5-fold lower affinity (J.P. DiSanto et al. Proc. Natl. Acad. Sci. 1994, 91:9466-9470). These isoforms produce an atypical form of an X chromosome-linked severe combined immunodeficiency disease.

The presence of splice variants can be used as diagnostic markers of diseases associated with genetic mutations. For example, the expression of the exon 6 splice variant (v6) of the cell adhesion molecule CD44 is correlated with the expression of the tumor suppressor gene p53. Both have been shown to be markers of tumor progression in colorectal cancer (J.W. Mulder et al. Gut 1995, 36:76-80; Y. Matsumura Lancet 1992, 340:1053-1058). Asymptomatic carriers of the acute intermittent porphyria were identified by identification of a mutant allele containing a CG to CT transversion at the exon1/intron 1 boundary via in vitro amplification of DNA followed by hybridization of the target sequence to allele-specific oligonucleotides.

Accordingly, splicing variants have been observed in several gene loci and several diseases. Identification of thes variants has proven to be especially useful in diagnosis and detection of asymptomatic carriers.

SUMMARY OF THE INVENTION

A novel insertional motif that arises from splice mutations or alternative utilization of cryptic or less preferred splic donor sites has now been identified. These splicing variations result in the in-frame insertion within a normal protein sequence of four amino acids, valine-arginine-X-glutamine (VRXQ), where X is a hydrophilic amino acid. This motif has been identified in splice variants of a receptor, an enzyme, and a putative channel protein, all of which are involved in normal neurological functioning. Identification of this motif allows for screening of genes and gene products for splice variations.

A method for the detection of this motif in expressed proteins in vitro or in situ with the use of specific antisera, polyclonal or monoclonal antibodies is provided. A method for the detection of allele-specific genetic mutations using selected oligonucleotides with standard hybridization-based detection techniques is also provided. A method for diagnosing Alzheimer's Disease (AD) by detecting differences in levels of transcripts having the VRXQ insertion or proteins encoded therefrom is further provided. A preferred embodiment of such method for detecting AD provides for the detection of Familial Adult Onset Alzheimer's Disease (FAD).

BRIEF DESCRIPTION OF THE FIGURES

Figure 1 is a schematic of potential alternative splicing with 3 exons and 4 introns.

Figure 2 is a schematic of the consensus exon-intron-exon structure and sequence.

Figure 3 provides the sequence of the VRSQ variant of the presenilin 1 gene. SEQ ID NOS: 1 - 2

Figure 4 provides PS-1 Oligonucleotide Probes. SEQ ID NOS: 3 - 5

Figure 5 provides tabulated results of quantification of the ISH signal for PS-1-long and PS-1-short mRNAs in human brain.

DETAILED DESCRIPTION OF THE INVENTION

The presenilin 1 (herein "PS-1") gene encodes a neuropeptide predicted to be a classical seven transmembran protein (Sherrington et al. Nature 1995, 375:754-760). Missense mutations within this gene have been found in several families exhibiting early-onset Alzheimer's disease. Genomic analysis has revealed the intron-exon boundaries of the hnRNA. A common polymorphism located within the intron 3' to exon 9 was identified in early onset AD patients. This polymorphism also showed a strong association with the occurrence of typical late onset AD families. This particular mutation did not produce an alteration in the coding sequence but is typical of variations leading to alternatively spliced proteins.

Other mutations within different introns of the PS-1 gene have been identified. These lead to alternatively spliced variants as well. One novel variant of the PS-1 protein isolated from a human cerebellar cDNA library contains a four amino acid insertion between codons 26 and 27 (VRSQ) (Figure 3). This variant arises from alternative use of a 5' exon donor site in the exon 3/intron 3 boundary and results in the loss of some potential phosphorylation sites. A similar motif (VRXQ- where X is a hydrophilic amino acid) arising from aberrant splicing has also arisen due to alternative splicing in several other neurological proteins as well.

For xample, th mRNA for tyrosin hydroxylase, the rate limiting nzym in the synthesis of catecholomines, can undergo alt rnativ splicing to produc several differ nt isoforms (Kobayashi et al. J. Biochem. 1988, 103(6) 907-12; Lewis et al. Neuroscience 1993, 54(2) 477-92). The id ntified variants contain a 12 bp insertion encoding the sequ nce VRGQ. Isoforms containing the VRGQ insertion have also been found to exhibit alterations in phosphorylation by MAP kinase (Sutherland et al. Eur J Biochem. 1993, 217(2) 715-22). Furthermore, a tyrosine hydroxylase variant containing this insertion has been implicated in Parkinson's disease.

Another neuropeptide, gamma-Aminobutyric acid A (GABAA) receptor, undergoes alternative splicing to yield a

multiplicity of transcripts (Whiting et al. P.N.A.S. 1990, 87(24) 9966-70; Lasham et al. Biochem. Soc. Trans. 1991, 19 (1) 9S). GABA receptors are multisubunit ligand gated ion channels which mediate neuronal inhibition by GABAA and are composed of at least four subunit types (alpha, beta, gamma, and delta). The beta 4 subunit can undergo alternative splicing at two 5'-donor splice sites separated by 12 bp in the region that encodes the presumed intracellular loop b tween transmembrane domains M3 and M4. The insertion of the 12 bp sequence results in the addition of a VREQ motif (Bateson et al. J. Neurochem 1991, 56(4) 1437-40).

In all three neurological proteins, the alternative splice site generates variants containing a specific motif (VRXQ) which appears to be intracellularly located and alters phosphorylation by various kinases.

In the present invention, a method for detecting the presence of the VRXQ motif in polyadenylated messenger RNA transcripts (polyA mRNA) and resultant expressed proteins, (where V is valine, R is arginine, X is any hydrophilic amino acid residue, and Q is glutamine) or in cDNA resulting from these RNAs is provided. A method for quantitating such transcripts encoding and proteins having a VRXQ motif are also provided. Oligonucleotides having the anticodon sequences associated with the VRXQ motif having degenerate positions at the third base position of each codon can be used for the detection and quantitation of mRNA. Additionally, these oligonucleotides can be associated with codon sequences and used for the detection of cDNAs, and quantitation of the transcript from which the cDNA was derived. For example, codon and anticodon oligonucleotides for VRNQ comprise GU(N) AG(A/G) AA(C/U) CA(A/G) and the reverse complement. Hybridization of appropriate oligonucleotides can be detected and quantitated directly by procedures well known to those of skill in the art using radioactively or fluorescently labeled oligonucleotides. Indirect detection and quantitation procedures such as, but not limited to, biotinylated oligonucleotides/strepavidin-horseradish peroxidase, enhanced chemiluminescent detection, or fluorescently tagged strepavidins can also be performed.

Specific antibodies against the VRXQ motif can also be used for detection of the motif and quantitation of proteins having the motif. Various procedures known in the art may be used for the production of such antibodies.

For example, these antibodies can be obtained by direct injection of a polypeptide containing a VRXQ motif into an animal, preferably a nonhuman. The antibody so obtained will then bind to polypeptides containing this motif. Such antibodies can then be used to isolate and quantitate polypeptides containing this motif from tissues.

For preparation of monoclonal antibodies, any technique which provides antibodies produced by continuous cell line cultures can be used. Examples include the hybridoma technique (Kohler and Milstein, Nature 1975, 256:495-497), the trioma technique, the human B-cell hybridoma technique (Kozbor et al., Immunology Today 1983, 4:72), and th EBV-hybridoma technique to produce human monoclonal antibodies (Cole et al. in Monoclonal Antibodies and Cancer Therapy, Alan R. Liss, Inc., 1985, pp. 77-96).

Techniques described for the production of single chain antibodies (U.S. Patent 4,946,778) can be adapted to produce single chain antibodies to the immunogenic motif of this invention. Also, transgenic mice may be used to express humanized antibodies to polypeptides containing this motif.

Primary antibody-antigen reactions can be visualized and quantitated secondarily by standard enzyme-linked immunosorbent assay (ELISA) procedures. An ELISA assay initially comprises preparing an antibody specific to a VRXQ motif, preferably a monoclonal antibody. In addition a reporter antibody is prepared against the monoclonal antibody. To the reporter antibody is attached a detectable reagent such as horse radish peroxidase. A sample is then removed from a host and incubated on a solid support, e.g., a polystyrene dish, that binds the proteins in the sample. Any fre protein binding sites on the dish are then covered by incubating with a non-specific protein like BSA. Next, the monoclonal antibody is incubated in the dish during which time the monoclonal antibodies attach to any proteins containing the VRXQ motif attached to the polystyrene dish. All unbound monoclonal antibody is washed out with buffer. Th reporter antibody linked to horseradish peroxidase is then placed in the dish resulting in binding of the reporter antibody to any monoclonal antibody bound to proteins containing the VRXQ motif. Unattached reporter antibody is then washed out. Peroxidase substrates are then added to the dish and the amount of color developed in a given time period is a measurement of the amount of protein containing the VRXQ motif present in a given volume of patient sample when compared against a standard curve to detect and quantitate the protein. Examples of other detectable reagents which can be used include, but are not limited to, luciferase and fluorescently or radioactively tagged secondary antibodies. Specific populations of immune cells or chimeric cells (e.g., hybridomas) that express antibodies to VRXQ epitopes on their cell surfaces and respond by degranulation or release of cellular contents such as histamines that can be detected functionally or preloaded radiolabeled metals such as chromium are also useful.

Embodiments of the invention can be used to detect alterations in and make comparisons between expression in of PS-1 variants in presumptive neurodegenerativ disease, particularly neurodegenerative dis ase associated with h ad injury and AD, and more particularly chromosom 14 FAD. In a particularly preferred embodim nt, probes and m thods of the invention can be used to d tect a reduction in the expression of PS-1 transcript encoding the VRSQ motif, shown by this invention to be a diagnostic marker for chromosome 14 FAD, since lowered levels are associat d with chromosome 14 FAD. Preferred embodiments of the invention provid for comparisons betw en variants comprising the VRSQ region with those lacking it enabling the diagnosis of AD, particularly chromosome 14 FAD.

The methods of the invention to detect and quantitate PS-1 polynucleotide sequence, PS-1 expression levels and

gene expr ssion products, particularly the immunological methods and methods using oligonucleotides, can be used with bodily tissues and fluids from individuals. Preferred bodily tissues and fluids useful with the methods of th invention include, but are not limited to, blood cells, plasma, skin cells, and brain cells, particularly neuronal, glial, and astrocyte cells.

The following examples are provided for illustrative purposes only and are not int nded to limit the invention.

EXAMPLES

Example 1

A novel splice variant of the PS-1 gene described by Sherrington et al. Nature 1995, 375:754-760, was isolated from a human cerebellar and a human fibroblast library. In this novel splice variant there is a deletion of four amino acids at codons 26-27 (VRSQ). This arises from alternative use of a 5' exon donor site in the exon3/intron 3 (-52 to 75 nt) boundary. The ...CAG/gta... boundary of the final Gln codon of exon 3 of the VRSQ motif provides a 5' exon AG donor site and GT intron consensus 5' boundary and use of this splice site results in the insertion of the 12-nts encoding the VRSQ motif. The upstream ...ACT/GTA... boundary of the Thr-Val codons provides the less preferred CT (AG preferred) 5' exonic boundary to the consensus GT 5' intronic boundary and splicing at this site would remove the VRSQ motif. Interestingly, in the PS-1 protein of Sherrington et al. Nature 1995, 375:754-760, this is the sole observed product and point mutations are interspersed elsewhere.

Example 2

In the GABA receptor 4 subunit alternative splicing adds a VREQ motif (Bateson et al. J. Neurochem 1991, 56(4) 1437-40). A chicken genomic cDNA library was screened with chicken beta- 4' subunit cDNA at high stringency. Southern blot analysis, using cDNA sequence specific oligonucleotides as probes and subsequent restriction mapping allowed the identification of overlapping DNA fragments containing the coding regions of the beta-4 subunit gene. These fragments were subcloned into pBluescript and sequenced. Complete sequencing of one of the clones revealed the presence of 12 bp in the part encoding the intracellular loop (amino acid residues 335-338). Analysis of the beta-4 subunit gene reveals that the different transcripts encoding the two variants (absence or presence of 12bp loop) arise by the use of one of two 5'-donor splice sites (located in the intron immediately 3' of the 12 bp sequence).

Example 3

The expression of two PS-1 mRNA transcripts, one containing (herein " PS-1-long ") and one lacking the VSRQ motif (herein "PS- 1-short"), in the brains of patients with early onset FAD was analyzed. In situ hybridization (ISH) was used to determine the qualitative and quantitative pattern of expression of PS-1 mRNA in the brains of early onset (presumptive chromosome 14-linked) FAD cases; comparisons with brains from patients with late onset AD and from normal individuals were made.

In Situ Hybridization

PS-1 mRNA expression was examined in 4 neurologically normal control cases, 6 late onset AD cases and 3 early onset FAD cases. The late onset cases were thought to be of a sporadic nature as there was no evidence of family history and the mean age at death was 81.2 years (range: 79-84 years); they had a mean post mortem delay of 8.3 hours. The early onset FAD cases were presumed to be linked to chromosome 14 as they all had onset ages, family history, clinical presentations and histopathology typical of chromosome 14-linked FAD. For these the mean age at death was 45 years (range: 44-46 years) and the mean post mortem delay was 41.7 hours. All AD cases were diagnosed according to standard pathological criteria (Khachaturian, 1985, Archives of Neurology. 42:1097-1105). The controls had a mean age at death of 68.8 years (range: 57-85 years) and mean post mortem delay of 11.8 hours. The brain regions examined were the hippocampus, temporal cortex and frontal cortex (regions severely affected by AD pathology), the visual cortex (an area relatively unaffected, but which at the time of death may be in the early stages of the disease proc ss) and the cerebellum (an area not affected by th classic pathology associated with AD and with no clinical involv ment).

Three different oligoprobes were chosen and synth sized (Figur 4): one to d tect PS-1-long, one to PS-1-short and one that recognizes both transcripts, PS-1-both, Thes probes are not prėdicted to det ct the transcripts of pr senilin-2, a closely related gene on chromosome 1 (Rogaev, et al., 1995, Nature 376:775-78).

The ISH methodology is well known in the art and has been described in detail elsewhere (Najlerahim et al., 1990, FEBS Letters 7:317-333). For the ISH analyses 10(m cryostat tissue sections were used. Probes were labelled at their

3' end with ^{35}S-dATP using the NEN DuPont 3' end labelling system. Hybridization and wash temperatures for the various probes are given in Figure 4. Hybridized sections were apposed to tritium-sensitive film for the generation of autoradiographs. Hybridization with the PS-1 probes in the sense orientation on adjacent sections were used to control for non-specific background. The signal on autoradiographic film was quantified using an image analyzer (Seescan®). A representative area over most of a tissue section was measured: for example, in the hippocampus the different subfields were not separately quantified. The background signal (sense strand hybridization) was subtracted from the antisense signal. Statistical analysis of the data was performed using the well known two-tailed Student's t-test.

Northern Analysis

Northern analysis was carried out with the PS-1-both probe on a Northern blot (Clontech®, catalogue number: 7750-1) containing polyA+ mRNA from a number of different human brain regions. The probe was 3' end labelled with 32P-dATP using terminal transferase and hybridized under standard conditions (Clontech®, data sheet).

Diagnostic Methods and Reagents for FAD

In situ hybridization using all three probes revealed that PS-1 mRNA was present in all of the brain regions examined. Hybridization with a sense strand control probe gave a very low background signal. In the cerebral cortex (three regions) a signal was detected in both the grey and white matter, often with a similar intensity. A diffuse rather than laminar pattern was observed in grey matter and in the hippocampus the different subfields were not readily delineated (although the dentate gyrus was sometimes visible). In the cerebellum, the granule cell layer contained the most labelling. These data are consistent with PS-1 mRNA expression in both neurons and glia.

Northern analysis confirmed that the PS-1-both oligoprobe detected a major transcript in human brain of the correct size for PS-1 mRNA (in accordance with the sequence data of Sherrington et al, 1995, Nature 375:754-760). A major band of approximately 3.4 kb was detected in all brain regions examined, indicating a wide distribution in brain for PS-1 mRNA. The observation of PS-1 mRNA in corpus callosum is consistent with the interpretation from our ISH data that PS-1 is expressed in glia.

A similar anatomical pattern was seen by ISH, in each region, for both PS-1-long and PS-1-short transcripts. Nevertheless there appeared to be differences between the transcripts in their levels of expression according to brain region; for example PS-1-short was relatively less abundant in the cerebellum (Figure 5).

The hybridization pattern was similar for the controls, sporadic AD and FAD cases. Quantification of the autoradiographic film revealed a statistically significant reduction in the amount of PS-1-long mRNA in FAD hippocampus and frontal cortex compared with the sporadic AD cases (Figure 5; $p = 0.003$ and $p = 0.014$ respectively). In the cerebellum there was no significant difference between the controls, sporadic AD and FAD cases. The reduction in PS-1-long appears to be specific because there was no change in the level of expression of PS-1-short mRNA in any brain region investigated between the three different groups (Figure 5), which indicates reasonable data consistency.

SEQUENCE LISTING

(1) GENERAL INFORMATION:

(i) APPLICANT: University of South Florida, Washington University and SmithKline Beecham Corporation

(ii) TITLE OF INVENTION: A Novel Diagnostic Marker for Splicing Variants of Genes Associated with Neurological Function

(iii) NUMBER OF SEQUENCES: 5

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(v) COMPUTER READABLE FORM:

(A) MEDIUM TYPE: DISKETTE, 3.5 INCH, 1.44 Mb STORAGE

(B) COMPUTER: IBM 486

(C) OPERATING SYSTEM: WINDOWS FOR WORKGROUPS

(D) SOFTWARE: WORDPERFECT 5.1

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(2) INFORMATION FOR SEQ ID NO: 1:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1914

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 1:

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(2) INFORMATION FOR SEQ ID NO: 2:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 463

(B) TYPE: Amino Acid

(D) TOPOLOGY: Linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 2:

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ARG GLN GLU HIS ASN ASP ARG ARG SER LEU GLY HIS PRO GLU PRO
                35                  40                  45

LEU SER ASN GLY ARG PRO GLN GLY ASN SER ARG GLN VAL VAL GLU
                50                  55                  60

GLN ASP GLU GLU GLU ASP GLU GLU LEU THR LEU LYS TYR GLY ALA
                65                  70                  75

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                80                  85                  90

VAL VAL VAL ALA THR ILE LYS SER VAL SER PHE TYR THR ARG LYS
                95                  100                 105

ASP GLY GLN LEU ILE TYR THR PRO PHE THR GLU ASP THR GLU THR
                110                 115                 120

VAL GLY GLN ARG ALA LEU HIS SER ILE LEU ASN ALA ALA ILE MET
                125                 130                 135

ILE SER VAL ILE VAL VAL MET THR ILE LEU LEU VAL VAL LEU TYR
                140                 145                 150

LYS TYR ARG CYS TYR LYS VAL ILE HIS ALA TRP LEU ILE ILE SER
                155                 160                 165

SER LEU LEU LEU LEU PHE GLU GLU SER PHE ILE TYR LEU GLY GLU
                170                 175                 180
```

VAL PHE LYS THR TYR ASN VAL ALA VAL ASP TYR ILE THR VAL ALA
185 190 195

LEU LEU ILE TRP ASN PHE GLY VAL VAL GLY MET ILE SER ILE HIS
200 205 210

TRP LYS GLY PRO LEU ARG LEU GLN GLN ALA TYR LEU ILE MET ILE
215 220 225

SER ALA LEU MET ALA LEU VAL PHE ILE LYS TYR LEU PRO GLU TRP
230 235 240

THR ALA TRP LEU ILE LEU ALA VAL ILE SER VAL TYR ASP LEU VAL
245 250 255

ALA VAL LEU CYS PRO LYS GLY PRO LEU ARG MET LEU VAL GLU THR
260 265 270

ALA GLN GLU ARG ASP GLU THR LEU PHE PRO ALA LEU ILE TYR SER
275 280 285

SER THR MET VAL TRP LEU VAL ASN MET ALA GLU GLY ASP PRO GLU
290 295 300

ALA GLN ARG ARG VAL SER LYS ASN SER LYS TYR ASN ALA GLU SER
305 310 315

THR GLU ARG GLU SER GLN ASP THR VAL ALA GLU ASN ASP ASP GLY
320 325 330

GLY PHE SER GLU GLU TRP GLU ALA GLN ARG ASP SER HIS LEU GLY
335 340 345

PRO HIS ARG SER THR PRO GLU SER ARG ALA ALA VAL GLN GLU LEU
350 355 360

SER SER SER ILE LEU ALA GLY GLU ASP PRO GLU GLU ARG GLY VAL
365 370 375

LYS LEU GLY LEU GLY ASP PHE ILE PHE TYR SER VAL LEU VAL GLY
380 385 390

LYS ALA SER ALA THR ALA SER GLY ASP TRP ASN THR THR ILE ALA
395 400 405

CYS PHE VAL ALA ILE LEU ILE GLY LEU CYS LEU THR LEU LEU LEU
410 415 420

LEU ALA ILE PHE LYS LYS ALA LEU PRO ALA LEU PRO ILE SER ILE
425 430 435

THR PHE GLY LEU VAL PHE TYR PHE ALA THR ASP TYR LEU VAL GLN
440 445 450

PRO PHE MET ASP GLN LEU ALA PHE HIS GLN PHE TYR ILE
455 460

(2) INFORMATION FOR SEQ ID NO: 3:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 30 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: Other

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 3:

GCACTCAATT CTGAATGCTG CCATCATGAT 30

(2) INFORMATION FOR SEQ ID NO: 4:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 30 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: Other

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 4:

AGCAATACTG TACGTAGCCA GAATGACAAT 30

(2) INFORMATION FOR SEQ ID NO: 5:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 29 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: Other

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 5:

CACCTGAGCA ATACWATGAC AATAGAGAA 29

Possible Final mRNAs

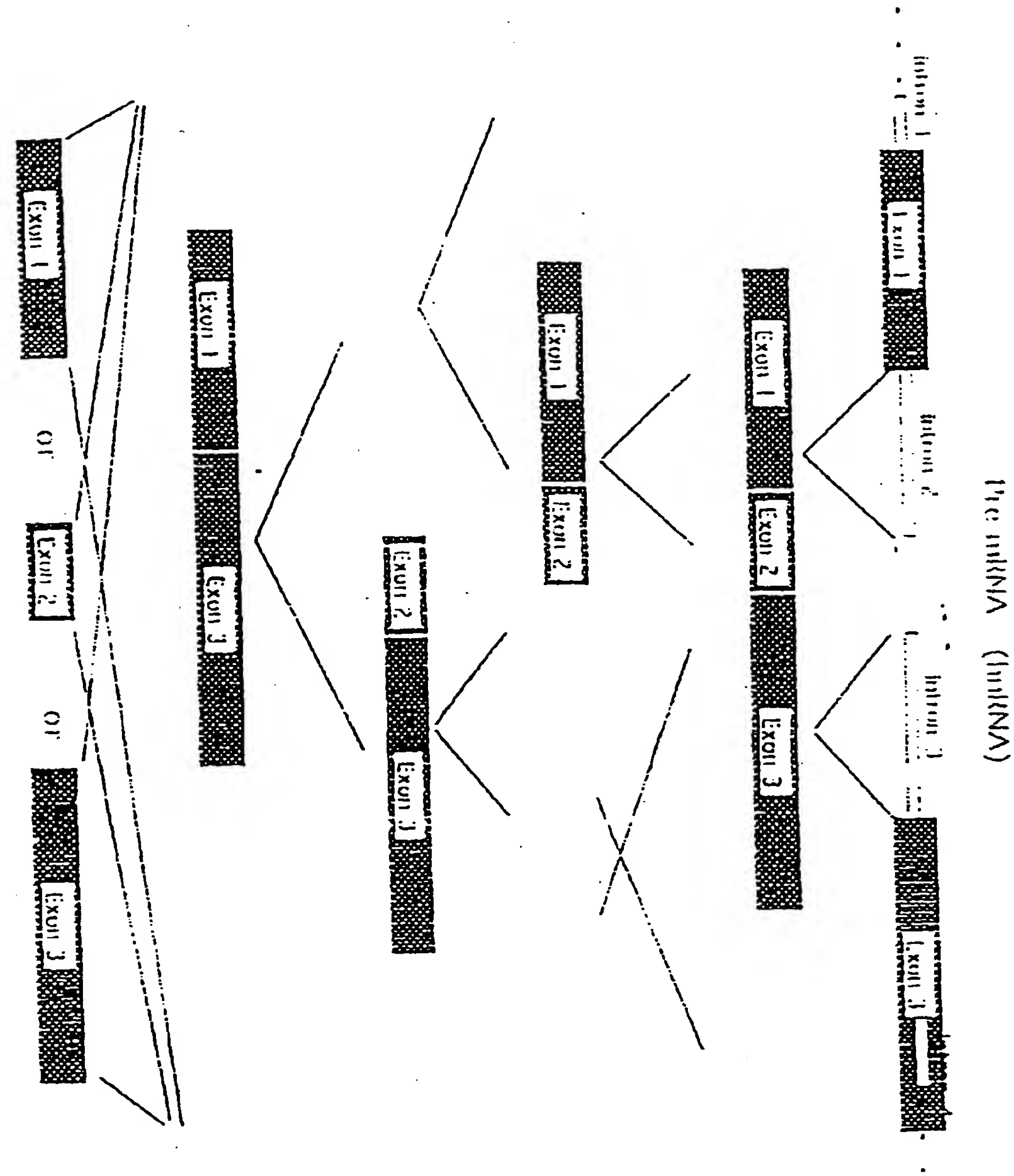

Figure 1

Structure of Exon-Intron Boundaries
and
Intron Consensus Sequences

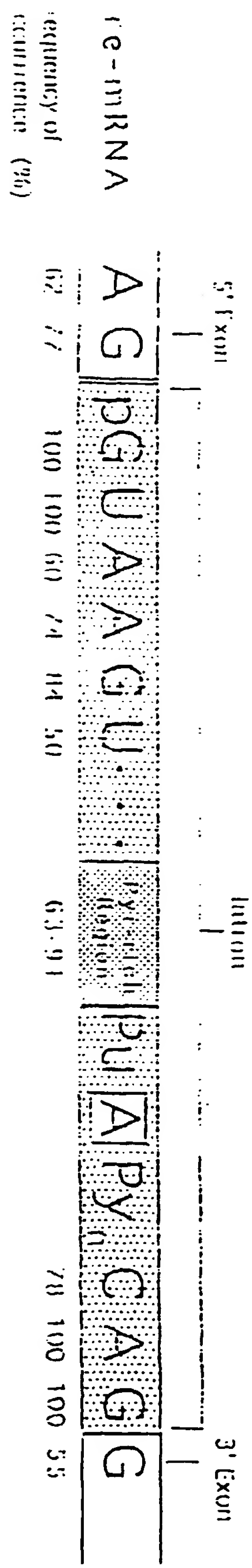

Figure 2

FIGURE 1A

```
                 11                  20                  29                  38                  47                  56
5' CCG TAC GTA GCC GCG GCG GCA GCG GGG CGG CGG GGA AGC GTA TGC ATA CAA ATT
   --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---

           65          74          83          92         101         110
   TAT TAG CAT GCA GAC TGG GAG AAC CAC AAG ACC TAA TCT CGG AGC CTG CAA GTG
   --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---

          119         128         137         146         155         164
   ACA ACA GCC TTT GCG GTC CTT AGA CAG CTT GGC CTG GAG GAG AAC ACA TGA AAG
   --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---

          173         182         191         200         209         218
   AAA GAA CCT CAA GAG GCT TTG TTT TCT GTG AAA CAG TAT TTC TAT ACA GTT GCT
   --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---

          227         236         245         254         263         272
   CCA ATG ACA GAG TTA CCT GCA CCG TTG TCC TAC TTC CAG AAT GCA CAG ATG TCT
   --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
        M   T   E   L   P   A   P   L   S   Y   F   Q   N   A   Q   M   S

          281         290         299         308         317         326
   GAG GAC AAC CAC CTG AGC AAT ACT AAT GAC AAT AGA GAA CGG CAG GAG CAC AAC
   --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
    E   D   N   H   L   S   N   T   N   D   N   R   E   R   Q   E   H   N

          335         344         353         362         371         380
   GAC AGA CGG AGC CTT GGC CAC CCT GAG CCA TTA TCT AAT GGA CGA CCC CAG GGT
   --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
    D   R   R   S   L   G   H   P   E   P   L   S   N   G   R   P   Q   G

          389         398         407         416         425         434
   AAC TCC CGG CAG GTG GTG GAG CAA GAT GAG GAA GAA GAT GAG GAG CTG ACA TTG
   --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
    N   S   R   Q   V   V   E   Q   D   E   E   E   D   E   E   L   T   L

          443         452         461         470         479         488
   AAA TAT GGC GCC AAG CAT GTG ATC ATG CTC TTT GTC CCT GTG ACT CTC TGC ATG
   --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
    K   Y   G   A   K   H   V   I   M   L   F   V   P   V   T   L   C   M

          497         506         515         524         533         542
   GTG GTG GTC GTG GCT ACC ATT AAG TCA GTC AGC TTT TAT ACC CGG AAG GAT GGG
   --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
    V   V   V   V   A   T   I   K   S   V   S   F   Y   T   R   K   D   G
```

FIGURE 1B

```
                551             560             569             578             587             596
CAG CTA ATC TAT ACC CCA TTC ACA GAA GAT ACC GAG ACT GTG GGC CAG AGA GCC
--- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
 Q   L   I   Y   T   P   F   T   E   D   T   E   T   V   G   Q   R   A

                605             614             623             632             641             650
CTG CAC TCA ATT CTG AAT GCT GCC ATC ATG ATC AGT GTC ATT GTT GTC ATG ACT
--- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
 L   H   S   I   L   N   A   A   I   M   I   S   V   I   V   V   M   T

                659             668             677             686             695             704
ATC CTC CTG GTG GTT CTG TAT AAA TAC AGG TGC TAT AAG GTC ATC CAT GCC TGG
--- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
 I   L   L   V   V   L   Y   K   Y   R   C   Y   K   V   I   H   A   W

                713             722             731             740             749             758
CTT ATT ATA TCA TCT CTA TTG TTG CTG TTC TTT TTT TCA TTC ATT TAC TTG GGG
--- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
 L   I   I   S   S   L   L   L   L   F   F   F   S   F   I   Y   L   G

                767             776             785             794             803             812
GAA GTG TTT AAA ACC TAT AAC GTT GCT GTG GAC TAC ATT ACT GTT GCA CTC CTG
--- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
 E   V   F   K   T   Y   N   V   A   V   D   Y   I   T   V   A   L   L

                821             830             839             848             857             866
ATC TGG AAT TTT GGT GTG GTG GGA ATG ATT TCC ATT CAC TGG AAA GGT CCA CTT
--- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
 I   W   N   F   G   V   V   G   M   I   S   I   H   W   K   G   P   L

                875             884             893             902             911             920
CGA CTC CAG CAG GCA TAT CTC ATT ATG ATT AGT GCC CTC ATG GCC CTG GTG TTT
--- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
 R   L   Q   Q   A   Y   L   I   M   I   S   A   L   M   A   L   V   F

                929             938             947             956             965             974
ATC AAG TAC CTC CCT GAA TGG ACT GCG TGG CTC ATC TTG GCT GTG ATT TCA GTA
--- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
 I   K   Y   L   P   E   W   T   A   W   L   I   L   A   V   I   S   V

                983             992            1001            1010            1019            1028
TAT GAT TTA GTG GCT GTT TTG TGT CCG AAA GGT CCA CTT CGT ATG CTG GTT GAA
--- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
 Y   D   L   V   A   V   L   C   P   K   G   P   L   R   M   L   V   E

               1037            1046            1055            1064            1073            1082
ACA GCT CAG GAG AGA GAT GAA ACG CTT TTT CCA GCT CTC ATT TAC TCC TCA ACA
--- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
 T   A   Q   E   R   D   E   T   L   F   P   A   L   I   Y   S   S   T

               1091            1100            1109            1118            1127            1136
ATG GTG TGG TTG GTG AAT ATG GCA GAA GGA GAC CCG GAA GCT CAA AGG AGA GTA
```

FIGURE 1C

```
--- --- --- --- --- --- -   --- --- --- --- --- --- --- ---   - --- ---
 M   V   W   L   V   N   M   A   E   G   D   P   E   A   Q   R   R   V

    1145        1154        1163        1172        1181        1190
TCC AAA AAT TCC AAG TAT AAT GCA GAA AGC ACA GAA AGG GAG TCA CAA GAC ACT
--- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
 S   K   N   S   K   Y   N   A   E   S   T   E   R   E   S   Q   D   T

    1199        1208        1217        1226        1235        1244
GTT GCA GAG AAT GAT GAT GGC GGG TTC AGT GAG GAA TGG GAA GCC CAG AGG GAC
--- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
 V   A   E   N   D   D   G   G   F   S   E   E   W   E   A   Q   R   D

    1253        1262        1271        1280        1289        1298
AGT CAT CTA GGG CCT CAT CGC TCT ACA CCT GAG TCA CGA GCT GCT GTC CAG GAA
--- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
 S   H   L   G   P   H   R   S   T   P   E   S   R   A   A   V   Q   E

    1307        1316        1325        1334        1343        1352
CTT TCC AGC AGT ATC CTC GCT GGT GAA GAC CCA GAG GAA AGG GGA GTA AAA CTT
--- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
 L   S   S   S   I   L   A   G   E   D   P   E   E   R   G   V   K   L

    1361        1370        1379        1388        1397        1406
GGA TTG GGA GAT TTC ATT TTC TAC AGT GTT CTG GTT GGT AAA GCC TCA GCA ACA
--- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
 G   L   G   D   F   I   F   Y   S   V   L   V   G   K   A   S   A   T

    1415        1424        1433        1442        1451        1460
GCC AGT GGA GAC TGG AAC ACA ACC ATA GCC TGT TTC GTA GCC ATA TTA ATT GGT
--- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
 A   S   G   D   W   N   T   T   I   A   C   F   V   A   I   L   I   G

    1469        1478        1487        1496        1505        1514
TTG TGC CTT ACA TTA TTA CTC CTT GCC ATT TTC AAG AAA GCA TTG CCA GCT CTT
--- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
 L   C   L   T   L   L   L   L   A   I   F   K   K   A   L   P   A   L

    1523        1532        1541        1550        1559        1568
CCA ATC TCC ATC ACC TTT GGG CTT GTT TTC TAC TTT GCC ACA GAT TAT CTT GTA
--- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
 P   I   S   I   T   F   G   L   V   F   Y   F   A   T   D   Y   L   V

    1577        1586        1595        1604        1613        1622
CAG CCT TTT ATG GAC CAA TTA GCA TTC CAT CAA TTT TAT ATC TAG CAT ATT TGC
--- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
 Q   P   F   M   D   Q   L   A   F   H   Q   F   Y   I   *

    1631        1640        1649        1658        1667        1676
GGT TAG AAT CCC ATG GAT GTT TCT TCT TTG ACT ATA ACA AAA TCT GGG GAG GAC
--- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---
```

Figure 5.

Quantification of the ISH signal for PS-1-long and PS-1-short mRNAs in human brain.

Brain region	Case	PS-1-long (n)	PS-1-short (n)
Hippocampus	Control	0.025 ± 0.014 (2)	0.023 (1)
	AD	0.035 ± 0.007 (3)	0.026 ± 0.01 (3)
	FAD	0.008 ± 0.001 (3)*	0.030 ± 0.004 (3)
Frontal cortex	AD	0.024 ± 0.005 (3)	0.042 ± 0.014 (3)
	FAD	0.012 ± 0.0 (3)**	0.022 ± 0.011 (3)
Cerebellum	Control	0.036 (1)	0.013 (1)
	AD	0.024 ± 0.007 (3)	0.019 ± 0.006 (3)
	FAD	0.012 ± 0.002 (2)	0.014 ± 0.005 (2)
Temporal cortex	FAD	0.014 ± 0.009 (3)	0.015 ± 0.01 (3)
Visual Cortex	FAD	0.016 ± 0.007 (3)	0.032 ± 0.001 (3)

values represent means ± s.d.; units are arbitrary (machine grey levels). *FAD vs AD p = 0.003; **FAD vs AD p = 0.014; Student's t-test.

Claims

1. A method of identifying an individual susceptible to a neurological disease comprising:

   providing a sample of genetic material from an individual susceptible to a neurological disease; and detecting the presence of an alternative splice site comprising the sequence VRXQ, wherein V is valine, R is arginine, X is any hydrophobic amino acid residue and Q is glutamine, in a polyadenylated messenger RNA transcript or protein encoded therefrom in the sample of g n tic mat rial.

2. The method of claim 1 wherein the sequence VRXQ is detected using selected oligonucleotide probes comprising anticodon sequences associated with the sequence VRXQ having degenerate positions at the third base position.

3. The m thod of claim 2 further comprising associating said oligonucleotides with codon sequences and detecting cDNA.

FIGURE 1D

```
        1685        1694        1703        1712        1721        1730
AAA GGT GRT TTT CCT GTG TCC CAC ATC TAA CAA AGT CAA GAT TCC CGK CTG GAC
--- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---

        1739        1748        1757        1766        1775        1784
TTT TGC AGC TTC CTK CCA AGT CTT CCT GAC CAC CTT GCA CTW TTG GAC TTT GGA
--- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---

        1793        1802        1811        1820        1829        1838
RGG AGG TGC CTA KAG AAA ACG RTT TTG AMC ATA CTT CAT CGC AGT GGA CTG TGT
--- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---

        1847        1856        1865        1874        1883        1892
CCC TCG GTG CAG AAA CTA CCA GAT TTG AGG GAC GAG GTC AAG GAG ATA TGA TAG
--- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---

        1901        1910
GCC CGG AAG TTG CTG TGC CCA TCA 3'
--- --- --- --- --- --- --- ---
```

Figure 4

PS-1 Oligonucleotide Probes

Probe	Sense Sequence	Bases *	Ti °C	Tw °C
PS-1-both	5'-GCACTCAATTCTGAATGCTGCCATCATGAT-3' SEQ ID NO: 3	638-667	24	50
PS-1-long	5-'AGCAATACT<u>GTACGTAGCCAG</u>AATGACAAT-3' SEQ ID NO: 4	315-344	23	49
PS-1-short	5'-CACCTGAGCAATACT/AATGACAATAGAGAA-3' SEQ ID NO: 5	309-323 and 336-350	22	47

*Refers to EMBL and GenBank entry HUMS182R (accession number: L42110); Sherrington et al 1995, Nature 375:754-760. Ti represents the hybridization temperature (incubation) and Tw represents the wash temperature. The underlined bases code for the amino acids V, R, S and Q.

4. The method of claim 1 wherein the sequence VRXQ is detected using an antibody against a polypeptide comprising the sequence VRXQ.

5. The method of claim 1 wherein the neurological disease comprises Alzheimer's Disease and the mRNA or protein is encoded by the presenilin 1 gene.

6. The method of claim 5 therein the sequence comprises a 4 amino acid insertion between codons 26 and 27 of the gene and the sequence VRSQ.

7. The method of claim 1 wherein the mRNA or protein is encoded by the gamma-Aminobutyric acid A receptor gene and the sequence comprises VREQ.

8. The method of claim 1 wherein the mRNA or protein is encoded by the tyrosine hydroxylase gene and the sequence comprises VRGQ.

9. A method for diagnosing a neurological disease comprising determining the levels of polyadenylated messenger RNA transcripts or proteins encoded therefrom comprising the sequence VRXQ wherein V is valine, R is arginine, X is any hydrophobic amino acid residue and Q is glutamine, in a sample of genetic material and comparing these levels with established controls.

10. The method of claim 9 wherein the neurological disease comprises Alzheimer's Disease and the mRNA or protein is encodes by the presenilin 1 gene.

11. The method of claim 10 wherein the sequence comprises a 4 amino acid insertion between codons 26 and 27 of the gene and the sequence VRSQ.

12. The method of claim 9 wherein the mRNA or protein is encoded by the gamma-Aminobutyric acid A receptor gen and the sequence comprises VREQ.

13. The method of claim 9 wherein the mRNA or protein is encoded by the tyrosine hydroxylase gene and the sequenc comprises VRGQ.

European Patent Office

EUROPEAN SEARCH REPORT

Application Number

EP 97300988.9

DOCUMENTS CONSIDERED TO BE RELEVANT			
Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim	CLASSIFICATION OF THE APPLICATION (Int. Cl. 6)
A,D	NATURE GENETICS, vol. 8, no. 1, October 1994 S.G. KALER et al. "Occipital horn syndrome and a mild Menkes phenotype associated with splice site mutations at the MNK locus" pages 195-202 * Totality * --	1,9	C 12 Q 1/68
A,D	PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCES OF THE UNITED STATES OF AMERICA, vol. 91, September 1994 J.P. DISANTO et al. "Defective human interleukin 2 receptor gamma chain in an typical X chromosome-linked severe combined immunodeficiency with peripheral T cells" pages 9466-9470 * Totality * --	1,9	TECHNICAL FIELDS SEARCHED (Int. Cl.6)
A,D	BIOCHEMICAL AND BIOPHYSICAL RESEARCH COMMUNICATIONS, vol. 202, no. 1, 1994 T. HAMAGUCHI et al. "A new variant of muscle phosphofructokinase deficiency in a Japanese case with abnormal RNA splicing" pages 444-449 * Totality * --	1,9	C 12 Q
A	WO - A - 94/18 319 (INSTITUT NATIONAL DE LA SANTE ET DE LA RECHERCHE MEDICALE) * Claims 1-6,12-15 * ----	1,9	

The present search report has been drawn up for all claims

Place of search	Date of completion of the search	Examiner
VIENNA	02-06-1997	WOLF

CATEGORY OF CITED DOCUMENTS

X : particularly relevant if taken alone
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